

# Carbon tetrachloride-induced changes in the activity of phase II drug-metabolizing enzyme in the liver of male rats: role of antioxidants

S.A. Sheweita <sup>a,\*</sup>, M. Abd El-Gabar <sup>b</sup>, M. Bastawy <sup>b</sup>

<sup>a</sup> *Department of Bioscience and Technology, Institute of Graduate Studies and Research, Alexandria University, P.O. Box 832, 163 Horreya Avenue, Alexandria, Egypt*

<sup>b</sup> *Department of Chemistry, Faculty of Science, Cairo University, Beni Suef-Branch, Beni Suef, Egypt*

Received 1 March 2001; accepted 4 June 2001

## Abstract

Glutathione S-transferases and glutathione play an important role in the detoxification of most toxic agents. In the present study, the protective effects of some antioxidants (L-ascorbic acid (AA), vitamin E (VE) or garlic) on carbon tetrachloride-induced changes in the activity of alanine amino transferase (ALT), aspartate amino transferase (AST), glutathione S-transferase (GST), and the level of glutathione (GSH) and thiobarbituric acid reactive substances (TBARS) were studied. The activities of ALT, and AST were assayed in plasma, whereas the activity of GST and the levels of GSH and TBARS were determined in the livers of rats. The current study included two experiments. In the first experiment, animals received single oral dose of CCl<sub>4</sub> (400 mg/kg body weight) after administration of AA (100 mg/kg b.w.), VE (100 mg/kg b.w.) or garlic (800 mg/kg b.w.) as single oral doses. In the second experiment, rats received repeated oral doses of antioxidants for 12 consecutive days followed by a single oral dose of CCl<sub>4</sub> on the 13th day and killed after that by 24 h. Treatment of male rats with CCl<sub>4</sub> significantly increased the activity of ALT and AST in plasma, and the levels of both GSH and TBARS in the liver. On the other hand, CCl<sub>4</sub> inhibited the activity of GST after single dose treatment. Single-dose treatments of rats with AA, VE or garlic prior to the administration of CCl<sub>4</sub> could not restore the alterations in the activity of ALT and AST caused by CCl<sub>4</sub> to the normal control level. However, repeated dose treatments with these agents restored such alterations to the normal level. We observed that VE is more effective than AA and garlic in restoring the inhibition of GST activity caused by CCl<sub>4</sub> to the normal level after single dose treatments. On the other hand, AA and VE are more effective than garlic in restoring the induced TBARS level caused by CCl<sub>4</sub> to the normal control level after repeated dose treatments. It has been observed that the tested antioxidants were able to antagonize the toxic effects of CCl<sub>4</sub>, and such counteracting effects were more pronounced when they were administered as repeated doses prior to administration of CCl<sub>4</sub>. © 2001 Elsevier Science Ireland Ltd. All rights reserved.

**Keywords:** Antioxidants; Glutathione; Glutathione S-transferase; Free radicals

*Abbreviations:* AA, ascorbic acid; ALT, alanine amino transferase; AST, aspartate amino transferase; GSH, glutathione; GST, glutathione S-transferase; LDH, lactate dehydrogenase; TBARS, thiobarbituric acid reactive substances; VE, vitamin E.

\* Corresponding author. Tel.: +20-3-422-5007; fax: 20-3-428-5792.

E-mail address: ssheweita@yahoo.com (S.A. Sheweita).

## 1. Introduction

Four families of glutathione S-transferase termed alpha ( $\alpha$ ), mu ( $\mu$ ), pi ( $\pi$ ), and theta ( $\theta$ ) have been characterized. They have different but sometimes overlapping substrate specificities (Mannervik, 1985; Sato, 1989; Buetler, 1998). Conjugation of toxic metabolites with glutathione (GSH), catalyzed by glutathione S-transferases (GST), is one of the major pathways for the detoxification of toxic metabolites (Jakoby and Habig, 1980). Previous studies have shown that GSH and GST can reduce the covalent binding of epoxides of well-known carcinogens, e.g. benzo(a)pyrene and aflatoxin B<sub>1</sub>, to DNA and consequently reduce hepatocarcinogenesis caused by these compounds (Lotlikar et al., 1980; Malaveill et al., 1980; Gopalan et al., 1992, 1994). Several dietary compounds have been demonstrated to reduce gastrointestinal cancer rates in both human and animals through induction of GST activity. For example, sulforaphane, indole-3-carbinol, D-limonene and relafen induced GST $\alpha$  levels in small intestine and livers, GST $\mu$  levels in stomach and small intestine, GST $\pi$  levels in stomach, small and large intestines (Van Lieshout et al., 1998). The hepatic activity of GSTs and UDP-glucuronyl transferase were induced after administration of aqueous extracts of either green or black tea to rats as the sole drinking fluid for 4 weeks (Van Lieshout et al., 1998). The coffee specific diterpene cafestol and kahweol have been reported to be anti-carcinogenic in several animal models due to induction of glutathione S-transferase  $\pi$  class (Schilter et al., 1996). On the other hand, many different chemical compounds, parasites, and dietary items were found to inhibit the activity of GST in the livers of mice and also in human bladder tissues (Sheweita and Mostafa, 1996; Sheweita et al., 1997, 1998, 2001; Sheweita, 1998, 2000). In addition, animals treated with acriflavine, a protein kinase c inhibitor, and allyl disulfide showed complete blockage of GST gene expression as early as 12 h of treatment (Kim et al., 1998). Analogues of glutathione preferentially inhibit GST $\alpha$ , have less effect on  $\mu$  isozymes, and finally have little effect on rat  $\theta$  and  $\pi$  isozymes (Ebisawa and Deguchi, 1991; Ouwkerk-Mahadevan et al., 1995).

Carbon tetrachloride has been widely used to elicit experimental liver damage. CCl<sub>4</sub>-induced liver damage has been thought to depend on the formation of reactive intermediates such as trichloromethyl free radical produced by cytochrome P450 mixed function oxidase system (Recknagel et al., 1989), and further converted to a peroxy radical (Slater, 1987). These free radicals react with polyunsaturated fatty acids to propagate a chain reaction leading to lipid peroxidation or bind covalently to lipids and proteins, resulting in destruction of membranes (Slater, 1987; Recknagel et al., 1989). Peroxidation of the polyunsaturated fatty acids in biological membranes is believed to be the mechanism of toxicity for some chemicals such as carbon tetrachloride, ethanol, yellow phosphorus and orotic acid (DiLuzio, 1973; Recknagel et al., 1989). In addition to lipid peroxidation, carbon tetrachloride inhibited the activity of adenosine triphosphatase (ATPase), acetylcholine esterase, and glucose-6-phosphatase in rats (Gonzalez et al., 1996; El-Demerdash et al., 1999), and increased the activity of lactate dehydrogenase (El-Demerdash et al., 1999).

It has been reported that ascorbic acid and vitamin E act as antioxidants (Geetanjali et al., 1993; Guha et al., 1993; Meydani, 1995). Vitamin E is residing mainly in the cell membranes and thus helping to maintain membrane stability (Baker et al., 1996). The role of vitamin C as an antioxidant in the prevention and cure of diseases that result from free radicals has been of considerable interest and controversy lately (Barinaga, 1991). It minimizes the genotoxic effect not only of pesticides (Hoda and Sinha, 1991) but also other xenobiotics. Garlic (*Allium sativum*) is widely used as a part of diet and as a medicine (Al-Bekairi et al., 1990). In addition, garlic had a high antioxidant capacity against peroxy radicals (Cao et al., 1996) and contained active components lowering the levels of lipid and cholesterol (Myung et al., 1982). Considering the multifaceted role-played by vitamin C, vitamin E, and garlic, the present study was, therefore, conducted to investigate the changes in the activities of ALT, AST, GST, and the levels of GSH, and TBARS to evaluate the protective effect of these antioxidants against the toxicity of CCl<sub>4</sub>.

## 2. Materials and methods

### 2.1. Chemicals

Ascorbic acid (AA) as sodium salt, glutathione, bis-(3-carboxy-4-nitrophenyl)-disulfide, thiobarbituric acid, and 1-chloro-2,4-dinitrobenzene were obtained from Sigma Chemical Co. (St. Louis, MO, USA). Garlic powder was obtained from El-Nasr Company for Drying, Alexandria, Egypt. Vitamin E (DL  $\alpha$ -tocopherol acetate) was purchased from F. Hoffman–La Roche Ltd, Switzerland. Carbon tetrachloride was obtained from Merck, Germany.

### 2.2. Animals and administration schedule of CCl<sub>4</sub> and antioxidants

Male Sprague–Dawley rats weighing 130–150 g were used throughout the study. The local ethics committee approved the design of the experiments, and the protocol conforms to the guidelines of the NIH. Animals were housed seven animals per cage where food and water were provided ad libitum. Table 1 summarizes the protocol for administration of CCl<sub>4</sub> and antioxidants. Briefly, in single-dose experiment, carbon tetrachloride was administered directly as a single dose

after administration of AA, VE, or garlic (Table 1). In repeated-dose experiment, AA, VE, or garlic were administered for 12 consecutive days and CCl<sub>4</sub> was administered on the 13th day as a single dose. Rats in both single- and repeated-dose experiments were killed after 24 h of CCl<sub>4</sub> administration. The control animals received corn oil as vehicle. The number of animals in each treatment was five rats.

### 2.3. Enzymes assay

In both experiments, at the designated time point, animals were sacrificed by cervical dislocation. Freshly removed livers were washed and homogenized in 0.1 M potassium phosphate buffer, pH 7.6. The homogenates were centrifuged at 12000  $\times$  g for 20 min at 4 °C. The supernatants were used for estimation of glutathione S-transferase activity, glutathione levels, and thiobarbituric acid reactive substances. Glutathione (GSH) levels were estimated in the supernatant of liver tissue homogenates according to the method of Mitchell et al. (1973), using sulfosalicylic acid for protein precipitation and bis-(3-carboxy-4-nitrophenyl)-disulfide for color development. Glutathione S-transferase activity was assayed according to the method of Habig et

Table 1  
The protocol for administration of CCl<sub>4</sub> and antioxidants to male rats

Treatment	Experimental groups <sup>a</sup>															
	Single dose experiment (24 h) <sup>b</sup>								Repeated-dose experiment (12 consecutive days) <sup>c</sup>							
	1	2	3	4	5	6	7	8	1	2	3	4	5	6	7	8
Corn oil (1 ml/kg)	–	–	–	–	–	–	–	–	+	+	–	–	–	–	–	–
AA (mg/kg)	–	–	100	–	–	100	–	–	–	–	100	–	–	100	–	–
VE (mg/kg)	–	–	–	100	–	–	100	–	–	–	–	100	–	–	100	–
Garlic (mg/kg)	–	–	–	–	800	–	–	800	–	–	–	–	800	–	–	800
CCl <sub>4</sub> (mg/kg)	–	400	–	–	–	400	400	400	–	400	–	–	–	400	400	400

<sup>a</sup> The number of animals in each group was five rats.

<sup>b</sup> In single-dose experiment, groups 1, 2, 3, 4, and 5 received corn oil as vehicle, CCl<sub>4</sub>, AA, VE, and garlic respectively, whereas groups 6, 7, and 8 received a single dose of CCl<sub>4</sub> directly after administration of antioxidants.

<sup>c</sup> In repeated-dose experiment, groups 1, 3, 4, and 5 received corn oil as vehicle, AA, VE, and garlic respectively for 12 consecutive days, whereas groups 6, 7, and 8 received single dose of CCl<sub>4</sub> on the 13th day after administration of antioxidants. In addition, group 2 received single dose of CCl<sub>4</sub> on the 13th day. All animals group in either single- or repeated-dose experiments were killed after 24 h of CCl<sub>4</sub> administration.

al. (1974). The incubation mixture contained 30 µg protein of the liver supernatant fraction, 0.5 ml of glutathione (0.5 mM); 0.1 M sodium phosphate buffer, pH 7.3. After preincubation at 37 °C for 5 min the reaction was initiated by adding 50 µl of 1-chloro-2,4-dinitrobenzene (CDNB) (0.5 mM) in the incubation mixture and incubated at 37 °C for another 5 min. Trichloroacetic acid solution (0.2 ml, 33%) was added to terminate the reaction. After centrifugation, the CDNB conjugate was measured in the supernatant at 340 nm. Calculations were made using a molar extinction coefficient of  $9.6 \text{ mM}^{-1} \text{ cm}^{-1}$ . A unit of enzyme activity is defined as the amount of enzyme that catalyzes the formation of 1 µmol of CDNB conjugate/mg protein/min under the assay conditions. Thiobarbituric acid-reactive substances (TBARS) were measured as described by Tappel and Zalkin (1959). The color intensity of the reactants (TBARS) was measured at 532 nm. An extinction coefficient of  $156000 \text{ M}^{-1} \text{ cm}^{-1}$  was used for calculation. The protein concentration was measured by the method of Lowry et al. (1951), using bovine serum albumin as standard. Monitoring of serum amino transferases activity was carried out in the plasma according to the instructions of the commercially available kits (Boehringer Mannheim GmbH, Germany).

#### 2.4. Statistical analysis

Data analyses were conducted using General Linear Model Procedure of Statistical Analysis System (1995), and the difference between means was compared using least-squares difference (LSD) at 0.05 significant level (Steel and Torrie, 1980).

### 3. Results and discussion

Glutathione and glutathione S-transferase aid in the protection of cells from the lethal effects of toxic and carcinogenic compounds (Ketterer, 1988). Inducers of GSTs are generally considered as protective compounds against carcinogens. One of the hypotheses explaining the mechanism of chemopreventive activity of antioxidants against

carcinogens is that they activate detoxification system such as glutathione S-transferase (Hatono et al., 1996). The human diet contains many compounds that inhibit various steps of the carcinogenic process (Wattenberg, 1985; Nijhoff and Peters, 1994; Hatono et al., 1996). In agreement with these observations, single- and repeated-dose treatments of male rats with AA, VE or garlic induced the activity of glutathione S-transferase relative to  $\text{CCl}_4$ -treated group (Tables 2 and 3). This induction could be one of the protective mechanisms of AA, VE, or garlic against the toxic metabolites of  $\text{CCl}_4$ . Supporting this suggestion, it has been found that VE and AA reduced the toxicity and mortality caused in rats by cigarette smoking and aflatoxin B<sub>1</sub> (Chow et al., 1984; Netke et al., 1997). Also, consumption of garlic powder significantly decreased the *in vivo* binding of 7,12-dimethylbenzo(a)anthracene to mammary DNA and consequently depressed mammary tumors (Lin et al., 1992). Diallyl disulfide, a component of garlic oil, exerted an inhibitory effect in colon and renal carcinogenesis in rats treated with carcinogens (Takahashi et al., 1992). Moreover, ascorbic acid and  $\alpha$ -tocopherol acetate reduced the mutagenic effects of some heterocyclic amines (Edenharder et al., 1999).

On the other hand,  $\text{CCl}_4$  treatment as a single dose inhibited the activity of glutathione S-transferase in the livers of rats (Table 2). In agreement with the present study, it has been found that carbon tetrachloride and deoxycholic acid decreased the activity of GST in cultured rat hepatocytes (Lee et al., 1991). This inhibition could potentiate the toxicity of  $\text{CCl}_4$  upon the liver and probably other organs.

Glutathione is an important cellular factor influencing the effectiveness of a variety of chemotherapeutic and alkylating agents; organs with low GSH levels are more susceptible to the action of these agents, whereas those with high GSH levels are expected to be more protected and therefore resistant (Hamilton et al., 1985). In accordance with these observations, the mechanism of VE protection against the toxicity of  $\text{CCl}_4$  might be due to enhancing of GSH level since single- and repeated-dose treatments of rats with VE markedly induced the levels of GSH (Tables 2

Table 2

Influence of AA, VE or garlic after single-dose treatments prior to the administration of carbon tetrachloride on the activity of amino transferases (ALT and AST), glutathione S-transferase, and the levels of glutathione/TBARS in the liver of male rats

Parameters	Treatments								
	Control	CCl <sub>4</sub>	AA	VE	Garlic	AA+CCl <sub>4</sub>	VE+CCl <sub>4</sub>	Garlic+CCl <sub>4</sub>	
ALT	31.5 ± 0.48 <sup>E</sup>	131.5 ± 7.3 <sup>B</sup>	31.0 ± 2.35 <sup>E</sup>	61.5 ± 4.0 <sup>C</sup>	42.0 ± 1.9 <sup>D</sup>	153.0 ± 4.1 <sup>A</sup>	137.0 ± 1.5 <sup>B</sup>	135.0 ± 1.2 <sup>B</sup>	
AST	27.8 ± 3.0 <sup>CD</sup>	184.0 ± 13.0 <sup>B</sup>	21.5 ± 2.4 <sup>D</sup>	26.0 ± 3.4 <sup>D</sup>	51.7 ± 6.7 <sup>C</sup>	203.1 ± 13.2 <sup>AB</sup>	198.0 ± 9.5 <sup>B</sup>	226.0 ± 12.3 <sup>A</sup>	
GSH	8.7 ± 1.0 <sup>CD</sup>	13.5 ± 1.1 <sup>A</sup>	13.5 ± 0.8 <sup>A</sup>	14.3 ± 0.7 <sup>A</sup>	10.5 ± 1.0 <sup>BC</sup>	12.2 ± 0.6 <sup>AB</sup>	7.6 ± 0.54 <sup>D</sup>	9.0 ± 0.9 <sup>CD</sup>	
GST	0.7 ± 0.02 <sup>A</sup>	0.55 ± 0.01 <sup>CD</sup>	0.7 ± 0.01 <sup>AB</sup>	0.72 ± 0.05 <sup>A</sup>	0.6 ± 0.06 <sup>BC</sup>	0.57 ± 0.02 <sup>CD</sup>	0.65 ± 0.01 <sup>ABC</sup>	0.48 ± 0.02 <sup>D</sup>	
TBARS	0.150 ± 0.02 <sup>B</sup>	0.8 ± 0.021 <sup>E</sup>	0.14 ± 0.01 <sup>B</sup>	0.03 ± 0.002 <sup>D</sup>	0.1 ± 0.01 <sup>BC</sup>	0.22 ± 0.04 <sup>A</sup>	0.04 ± 0.01 <sup>CD</sup>	0.12 ± 0.02 <sup>B</sup>	

Note: means with different superscript letters differ significantly ( $P < 0.05$ ). Values are the mean ± SE of five male rats of each treatment. ALT, AST activity expressed as international unit (IU), which is defined as the amount of the enzyme that under defined assay conditions would catalyze 1 mol of substrate/min/liter. GSH value was expressed as μM GSH/g. liver. GST activity was expressed as unit/mg protein. TBARS level expressed as μM/g liver.

Table 3

Influence of AA, VE or garlic after repeated-dose treatments prior to the administration of carbon tetrachloride on the activity of amino transferases (ALT and AST), glutathione S-transferase, and the levels of glutathione/TBARS in the liver of male rats

Parameters	Treatments								
	Control	CCl <sub>4</sub>	AA	VE	Garlic	AA+CCl <sub>4</sub>	VE+CCl <sub>4</sub>	Garlic+CCl <sub>4</sub>	
ALT	38.5 ± 4.2 <sup>C</sup>	132.8 ± 4.6 <sup>A</sup>	33.3 ± 3.8 <sup>C</sup>	31.0 ± 2.2 <sup>C</sup>	28.0 ± 3.25 <sup>C</sup>	116.6 ± 9.9 <sup>B</sup>	43.0 ± 5.7 <sup>C</sup>	116.8 ± 6.0 <sup>B</sup>	
AST	15.5 ± 1.2 <sup>CDE</sup>	95.0 ± 10.3 <sup>A</sup>	18.1 ± 2.5 <sup>CD</sup>	16.7 ± 0.84 <sup>DE</sup>	15.0 ± 0.24 <sup>E</sup>	77.0 ± 5.2 <sup>B</sup>	12.6 ± 0.8 <sup>CDE</sup>	22.4 ± 3.0 <sup>F</sup>	
GSH	7.7 ± 0.6 <sup>C</sup>	13.5 ± 1.11 <sup>A</sup>	7.5 ± 0.3 <sup>C</sup>	9.6 ± 0.54 <sup>B</sup>	8.6 ± 0.3 <sup>BC</sup>	8.5 ± 0.6 <sup>BC</sup>	11.5 ± 0.7 <sup>A</sup>	8.2 ± 0.6 <sup>BC</sup>	
GST	0.8 ± 0.03 <sup>AB</sup>	0.5 ± 0.04 <sup>D</sup>	0.7 ± 0.03 <sup>C</sup>	0.9 ± 0.05 <sup>AB</sup>	0.7 ± 0.02 <sup>BC</sup>	0.6 ± 0.03 <sup>CD</sup>	0.9 ± 0.08 <sup>A</sup>	0.88 ± 0.1 <sup>AB</sup>	
TBARS	0.11 ± 0.03 <sup>CD</sup>	0.8 ± 0.08 <sup>A</sup>	0.1 ± 0.02 <sup>CD</sup>	0.04 ± 0.003 <sup>E</sup>	0.1 ± 0.01 <sup>CD</sup>	0.13 ± 0.009 <sup>CD</sup>	0.02 ± 0.003 <sup>E</sup>	0.2 ± 0.01 <sup>B</sup>	

Note: means with different superscript letters differ significantly ( $P < 0.05$ ). Values are the mean ± SE of five male rats of each treatment. ALT, AST activity expressed as international unit (IU), which is defined as the amount of the enzyme that under defined assay conditions would catalyze 1 mol of substrate/min/liter. GSH value was expressed as μM GSH/g. liver. GST activity was expressed as unit/mg protein. TBARS level expressed as μM/g liver.

and 3). In agreement with the present study, it has been found that red blood cells glutathione level was significantly greater in rats fed 1500 ppm vitamin E than rats fed 0 or 100 ppm vitamin E (Lii et al., 1998). GSH level did not change after repeated dose treatments of rats for 23 weeks with garlic (Khanum et al., 1998), and this is in agreement with the present study since repeated dose treatments of rats with garlic for 12 consecutive days did not change GSH level (Table 3). However, pretreatment of rats with garlic as single- or repeated-dose treatments restored the induced GSH levels caused by CCl<sub>4</sub> to the normal control levels (Tables 2 and 3). Also, pretreatment of rats with either single dose of VE and repeated dose of

AA restored the alteration in GSH to the normal level (Tables 2 and 3). Induction of GSH levels may be due to enhancing of GSH synthesizing enzymes such as γ-glutamylcysteine synthetase and GSH synthetase, which are key enzymes in the biosynthesis of glutathione (GSH) (Griffin, 1982; Seelig and Meister, 1985).

Carbon tetrachloride increased TBARS level after single-dose treatment (Table 2). Interestingly, pretreatment of rats with AA, VE or garlic as repeated doses prior to the administration of CCl<sub>4</sub> was found to reduce the induced level of TBARS caused by CCl<sub>4</sub> (Table 3). This recovery in TBARS level could be one of the protective mechanisms of AA, VE, or garlic against the

toxicity caused by  $\text{CCl}_4$  and probably other toxic compounds. Single- and repeated-dose treatments of rats with vitamin E only was found to reduce the level of free radicals (thiobarbituric acid-reactive substances) below the normal levels (Tables 2 and 3). Supporting our present study, it has been reported that vitamin E prevented lipid peroxidation through reduction of free radical level (Kawai-Kobayashi and Yoshida, 1986; Meydani, 1995; Lii et al., 1998) and also exerted a protective effect on  $\text{CCl}_4$ -induced hepatic fibrosis (Britton and Bacon, 1994).

Serum enzymes including aspartate amino transferase (AST) and alanine amino transferase (ALT) are used in the evaluation of hepatic disorders. An increase in these enzyme activities reflects active liver damage. Inflammatory hepatocellular disorders result in extremely elevated transaminase levels (Fortson et al., 1985; Hultcrantz et al., 1986). In accordance with these findings,  $\text{CCl}_4$  treatments has a significant role in the alteration of liver functions since the activity of AST and ALT were significantly higher than those of normal value (Table 2). In agreement with the present study, it has been found that treatment of mice with  $\text{CCl}_4$  induced the activity of lactate dehydrogenase (LDH) which can be used as a marker for hepatotoxicity (Animesh et al., 1997). Single-dose treatments with AA, VE or garlic could not recover the induction of AST or ALT activities caused by  $\text{CCl}_4$  to the normal control level (Table 2). However, repeated dose treatments of rats with VE or garlic restored the induction of AST caused by  $\text{CCl}_4$  to the normal control level (Table 3). Only vitamin E is effective in restoring the activity of ALT activity to the normal level (Table 3). It is obvious that VE is more effective than AA and garlic in recovering the alteration in ALT activity to the normal level after repeated dose treatments (Table 3).

Carbon tetrachloride-induced changes in the activity of glutathione S-transferase, amino transferases (ALT and AST) and also in the levels of reduced glutathione and free radicals. It is concluded that AA, VE or garlic substantially suppressed  $\text{CCl}_4$ -induced hepatotoxicity especially after repeated-dose treatments.

## References

- Al-Bekairi, A.M., Shah, A.H., Qureshi, S., 1990. Effect of *Allium sativum* on epididymal spermatozoa, estradiol-treated mice and general toxicity. *J. Ethnopharmacol.* 29, 117–125.
- Animesh, M., Anupam, B., Malay, C., Mandal, A., Bishayee, A., Chatterjee, M., 1997. *Trianthema portulacastrum* affords antihepatotoxic activity against carbon tetrachloride induced chronic liver damages in mice: reflection in subcellular level. *Phytother. Res.* 11, 216–221.
- Baker, H.W.G., Brindle, J., Irvine, D.S., Aitken, R.J., 1996. Protective effect of antioxidants on the impairment of sperm motility by activated polymorphonuclear leukocytes. *Fertil. Steril.* 65, 411–419.
- Barinaga, M., 1991. Vitamin C gets a little respect. *Science* 254, 374–376.
- Britton, R.S., Bacon, B.R., 1994. Role of free radicals in liver diseases and hepatic fibrosis. *Hepato-Gastroenterology* 414, 343–348.
- Buetler, T.M., 1998. Identification of glutathione S-transferase isozymes and gamma-glutamylcysteine synthetase as negative acute-phase proteins in rat liver. *Hepatology* 28 (6), 1551–1560.
- Cao, G., Sofic, E., Prior, R.L., 1996. Antioxidant capacity of tea and common vegetables. *J. Agric. Food Chem.* 44, 3426–3431.
- Chow, C.K., Chen, L.H., Thacker, R.P., Griffith, R.B., 1984. Dietary vitamin E and pulmonary biochemical responses of rats to cigarette smoking. *Environ. Res.* 34, 8–17.
- DiLuzio, N.R., 1973. Antioxidants, lipid peroxidation and chemical-induced liver injury. *Fed. Proc.* 32, 1875–1881.
- Ebisawa, T., Deguchi, T., 1991. Structure and restriction fragment length polymorphism of genes for human liver arylamine *N*-acetyltransferases. *Biochem. Biophys. Res. Commun.* 177 (3), 1252–1276.
- Edenharder, R., Worf-Wandelburg, A., Decker, M., Platt, K.L., 1999. Antimutagenic effects and possible mechanisms of action of vitamins and related compounds against genotoxic heterocyclic amines from cooked food. *Mutat. Res.* 444, 235–244.
- El-Demerdash, F.M., Yousef, M.I., Sheweita, S.A., 1999. Ameliorating effect of some antioxidants on carbon tetrachloride-induced-biochemical alterations in the brains of rats. *Environ. Nutr. Int.* 3, 245–255.
- Fortson, W.C., Tedesco, F.J., Starnes, E.C., Shaw, C.T., 1985. Marked elevation of serum transaminase activity associated with extrahepatic biliary tract disease. *J. Clin. Gastroenterol.* 76, 502–505.
- Geetanjali, D., Rita, P., Reddy, P.P., 1993. Effect of ascorbic acid in the detoxification of the insecticide dimethoate in the bone marrow erythrocytes of mice. *Food Chem. Toxicol.* 31, 435–437.
- Gonzalez, P.A., De-Toranzo, E.G.D., Castro, J.A., 1996. Depression of glucose 6-phosphatase activity in carbon tetrachloride-poisoned rats. *Free Radic. Biol. Med.* 21 (1), 81–87.

- Gopalan, P., Hirum, S., Lotlikar, P., 1994. Effect of glutathione levels on aflatoxin B1-DNA binding in livers and kidney of male rats and hamsters pretreated with buthionine sulfoximine and diethylmaleate. *Cancer Lett.* 76, 25–30.
- Gopalan, P., Jensen, D., Lotlikar, P., 1992. Glutathione conjugation of microsomes-mediated and synthetic aflatoxin B1-8,9-oxide by purified glutathione S-transferases from rats. *Cancer Lett.* 64, 225–233.
- Griffin, D.W., 1982. Mechanism of action, metabolism, and toxicity of buthionine sulfoximine and its higher homologues, potent inhibitors of glutathione synthesis. *J. Biol. Chem.* 257, 13704–13712.
- Guha, D., Dutta, K., Das, M., 1993. Vitamin C as antitoxic factor in DDT induced haematotoxicity in *Clarias batrachus*. *Proc. Zool. Soc. Calcutta* 46, 11–15.
- Habig, W., Pabst, M., Jakoby, W., 1974. Glutathione S-transferases: The first enzymatic step in mercapturic acid formation. *J. Biol. Chem.* 249, 7130–7139.
- Hamilton, T.C., Winker, M.A., Lovie, K.G., 1985. Augmentation of adriamycin, mephalan, and cisplatin cytotoxicity in drug-resistant and sensitive human ovarian carcinoma cell lines by buthionine sulfoximine, mediated glutathione depletion. *Biochem. Pharmacol.* 34, 2583–2589.
- Hatono, S., Jimenez, A., Wargovich, M.J., 1996. Chemopreventive effect of S-allylcysteine and its relationship to the detoxification enzyme glutathione S-transferase. *Carcinogenesis* 17 (5), 1041–1044.
- Hoda, M.Q., Sinha, S.P., 1991. Protective role of ascorbic acid and vitamin B-complex against pesticide-induced clastogeny in bonemarrow cells of mice. *Int. J. Vitam. Nutr. Res.* 61, 155–158.
- Hulterantz, R., Glaumann, H., Lindberg, G., Nilsson, L.H., 1986. Liver investigation in 149 asymptomatic patients with moderately elevated activities of serum aminotransferases. *Scand. J. Gastroenterol.* 21 (1), 109–113.
- Jakoby, W.B., Habig, W.H., 1980. Glutathione transferases. In: Jakoby, W.B. (Ed.), *Enzymatic Basis of Detoxification*. Academic Press, New York, pp. 63–94.
- Kawai-Kobayashi, K., Yoshida, A., 1986. Effect of dietary ascorbic acid and vitamin E on metabolic changes in rats and guinea pigs exposed to PCB. *J. Nutr.* 116, 98–106.
- Ketterer, B., 1988. Protective role of glutathione and glutathione S-transferases in mutagenesis and carcinogenesis. *Mutat. Res.* 202, 343–361.
- Khanum, F., Anilakumar, K.R., Sudarshanakrishna, K.R., Viswanathan, K.R., 1998. Effects of feeding fresh garlic and garlic oil on detoxifying enzymes and micronuclei formation in rats treated with azoxymethane. *Int. J. Vitam. Nutr. Res.* 68 (3), 208–213.
- Kim, S.G., Cho, J.Y., Chung, Y.S., Ahn, E.T., Lee, K.Y., Han, Y.B., 1998. Suppression of xenobiotic-metabolizing enzyme expression in rats by acriflavine, a protein kinase C inhibitor. Effects on epoxide hydrolase, glutathione S-transferases, and cytochromes P450. *Drug Metab. Dispos.* 26 (1), 66–72.
- Lee, E., Yasuhiro, M., Furukawa, Y., Simizu, H., Kariya, K., 1991. Selective release of glutathione transferase subunits from primary cultured rat hepatocytes by carbon tetrachloride and deoxycholic acid. *Toxicology* 67, 237–248.
- Lii, C.K., Ko, Y.J., Chiang, M.T., Sung, W.C., Chen, H.W., 1998. Effect of dietary vitamin E on antioxidant status and antioxidant enzyme activities in Sprague–Dawley rats. *Nutr. Cancer* 32 (2), 95–100.
- Lin, J., Lin, R.I., Milner, J.A., 1992. Inhibition of 7,12-dimethylbenz(a)anthracene-induced mammary tumors and DNA adducts by garlic powder. *Carcinogenesis* 13 (10), 1847–1851.
- Lotlikar, P., Insetta, S., Lyons, P., 1980. Inhibition of microsomes mediated binding of aflatoxin B1 to DNA by glutathione S-transferase. *Cancer Lett.* 9, 143–149.
- Lowry, O.H., Rosebrough, N.J., Farr, A.L., Randall, R.J., 1951. Protein measurement with the Folin Phenol Reagent. *J. Biol. Chem.* 193, 269–275.
- Malaveill, C., Brun, G., Bartsch, H., 1980. Effect of glutathione and uridine 5'-diphosphoglucuronic acid and mutagenesis by benzo(a)pyrene and aflatoxin B1 in the *Salmonella*/microsome assay. *Mutat. Res.* 83, 15–24.
- Mannervik, B., 1985. The isoenzymes of glutathione transferase. *Adv. Enzymol.* 57, 357–417.
- Meydani, M., 1995. Vitamin E. *Lancet* 345, 170–175.
- Mitchell, J.R., Jollow, D.J., Potter, W.Z., Davis, D.C., Gillette, J.R., Brodi, B.B., 1973. Acetaminophen-induced hepatic necrosis. I. Role of drug metabolism. *J. Pharmacol. Exp. Ther.* 187, 185–194.
- Myung, C., Eunsook, T.K., Troy, J.S., 1982. Effect of garlic on lipid metabolism in rats fed cholesterol or lard. *J. Nutr.* 112, 241–248.
- Netke, S.P., Roomi, M.W., Taso, C., Niedzwiecki, A., 1997. Ascorbic acid protects guinea pigs from acute aflatoxin toxicity. *Toxicol. Appl. Pharmacol.* 143, 429–435.
- Nijhoff, W.A., Peters, W.H., 1994. Quantification of induction of rat oesophageal, gastric and pancreatic glutathione and glutathione S-transferases by dietary anticarcinogens. *Carcinogenesis* 15 (9), 1769–1772.
- Ouwkerk-Mahadevan, S., Van Boom, J.H., Dreef-Tromp, M.C., Ploemen, J.H., Meyer, D.J., Mulder, G.J., 1995. Glutathione analogues as novel inhibitors of rat and human glutathione S-transferase isoenzymes, as well as of glutathione conjugation in isolated rat hepatocytes and in the rat in vivo. *Biochem. J.* 308, 283–290.
- Recknagel, R.O., Glende, E.A., Dolak, J.A., Waller, R.L., 1989. Mechanism of CCl<sub>4</sub> toxicity. *Pharmacol. Ther.* 43, 139–154.
- Sato, K., 1989. Glutathione transferases as markers of preneoplasia and neoplasia. *Adv. Cancer Res.* 52, 205–255.
- Schilter, B., Perrin, I., Cavin, C., Huggett, A.C., 1996. Placental glutathione S-transferase (GST-P) induction as a potential mechanism for the anti-carcinogenic effect of the coffee-specific components cafestol and kahweol. *Carcinogenesis* 17 (11), 2377–2384.
- Seelig, G.F., Meister, A., 1985. Glutathione biosynthesis;  $\gamma$ -glutamylcysteine synthetase from rat kidney. *Methods Enzymol.* 113, 379–390.

- Sheweita, S.A., Habib, S.L., Mostafa, M.H., 1997. Schistosomiasis induced changes in the glutathione levels and glutathione reductase/glutathione S-transferase in the human liver. *Biomed. Lett.* 56, 119–127.
- Sheweita, S.A., Mangoura, S.A., El-Shemi, A.G., 1998. Different levels of *Schistosoma mansoni* infection induce changes in drug-metabolizing enzymes. *J. Helminthol.* 72, 72–77.
- Sheweita, S.A., 1998. Heavy metal-induced changes in the glutathione levels and glutathione reductase/glutathione S-transferase in the liver of male mice. *Int. J. Toxicol.* 17 (4), 383–392.
- Sheweita, S.A., 2000. Drug-metabolizing enzymes: mechanisms and functions. A review. *Current Drug Metab.* 1 (2), 107–132.
- Sheweita, S.A., Abu El-Maati, M.R., El-Shahat, F.G., Bazeed, M.A., 2001. Changes in the expression of cytochrome P450 2E1 and the activity of carcinogen-metabolizing enzymes in *Schistosoma haematobium*-infected human bladder tissues. *Toxicology* 162 (1), 43–52.
- Sheweita, S.A., Mostafa, M.H., 1996. *N*-nitroso compounds and their effects on the hepatic level of glutathione, glutathione reductase and glutathione S-transferase activities in the liver of male mice. *Cancer Lett.* 99, 29–34.
- Slater, R.F., 1987. Free radicals and tissue injury: fact and fiction. *Br. J. Cancer* 8, 5–10.
- Statistical Analysis System, 1995. Principles and Procedure of Statistics, 2nd ed, McGraw-Hill, New York.
- Steel, R.G.D., Torrie, J.H., 1980. Principle and Procedure of Statistics. A Biochemical Approach, 2nd ed. McGraw-Hill, New York.
- Takahashi, S., Hakoi, K., Yada, H., Hirose, M., Ito, N., Fukushima, S., 1992. Enhancing effects of diallyl sulfide on hepatocarcinogenesis and inhibitory actions of the related diallyl disulfide on colon and renal carcinogenesis in rats. *Carcinogenesis* 13 (9), 1513–1518.
- Tappel, A.L., Zalkin, H., 1959. Inhibition of lipid peroxidation in mitochondria by vitamin E. *Arch. Biochem. Biophys.* 80, 333–336.
- Van Lieshout, E.M., Posner, G.H., Woodard, B.T., Peters, W.H., 1998. Effects of the sulforaphane analog compound 30:indole-3-carbinol, D-limonene or relafen on glutathione S-transferases and glutathione peroxidase of the rat digestive tract. *Biochim. Biophys. Acta* 1379 (3), 325–336.
- Wattenberg, L.W., 1985. Chemoprevention of cancer. *Cancer Res.* 45, 11–18.